





Blood 142 (2023) 458

## The 65th ASH Annual Meeting Abstracts

## ORAL ABSTRACTS

## 701.EXPERIMENTAL TRANSPLANTATION: BASIC AND TRANSLATIONAL

TCF-1is Required By CD8 + T Cells for the Maintenance of Alloimmune Responses in Graft-Vs-Host Disease Kevin Quann, MDPhD<sup>1</sup>, Faruk Sacirbegovic, PhD<sup>2</sup>, Sarah Rosenberger, MS<sup>1</sup>, Emily McFerran, BS<sup>1</sup>, Kentin Codispot, BS<sup>3</sup>, Warren Shlomchik, MD<sup>1</sup>

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Graft-vs-host disease (GVHD) is a common complication of allogeneic stem cell transplant (alloSCT) wherein donor T cells target alloantigens on recipient tissues. It is unclear how alloimmune responses are maintained in GVHD despite abundant antigen, which causes T cell anergy, deletion and exhaustion. Previously, we identified alloreactive TCF-1 high T cells arising post-transplant that resemble exhausted progenitors (T EXP) capable of propagating immune responses in other chronic antigen models. Here, we sought to further characterize these cells in the B6→129 MHC-matched GVHD mouse model, in which 129 recipients express the immunodominant H-2K b-restricted minor histocompatibility antigen (miHA) H60. At day +7 posttransplant, alloreactive CD8 + cells specific to H60 (as determined by MHC-I-tetramer staining; Tet H60+) were nearly uniformly PD-1 hiTox hi whereas Tet H60- cells displayed a bimodal distribution into discrete PD-1 hiTox hi and PD-1 loTox lo populations, indicative of more diverse antigen experiences. Among these both Tet H60+ and Tet H60- cells were TCF-1 hi cells. TCF-1 hi Tet H60+ cells were uniformly CD39 Tox hi PD-1 hi, which is a canonical T EXP phenotype. In contrast, among activated Tet H60- cells there were TCF-1 hi cells that were CD39 loTox hiPD-1 hi and Tox loPD-1 lo. At later times in spleen and lymph node, and in GVHD target tissues, these populations of TCF-1 + Tet H60+ and Tet H60- were found. To test if these CD39 10 TCF-1 hi T FXP had proliferative advantages in GVHD, we sorted congenic TCF-1 hiCD39 lo and TCF-1 loCD39 hi CD8 + cells from recipient spleens 14-days post-transplant and adoptively transferred them in competition in a 1:1 ratio (of Tet  $^{H60+}$  cells) into newly transplanted recipients. Among Tet H60+ cells in all tissues at day 14 post-transfer, TCF-1 hiCD39 lo-sorted progeny greatly outperformed TCF-1 loCD39 hi-sorted progeny. In line with their role as a source of GVHD effectors, progeny of TCF-1 hiCD39 lo cells were mostly TCF-1 loCD39 hi; however, a fraction remained TCF-1 hi consistent with their being able to undergo self-renewal. Conversely, we observed few if any TCF-1 + progeny of CD39 hi cells. We next tested whether TCF-1 was an important mediator of T cell fitness or whether it was only a marker for functionality. To do so we competed congenic wild-type (WT) and Tcf7 p45-/-(p45  $^{-1}$ ) donor CD8 cells, which lack the N-terminal  $\beta$ -catenin binding domain of TCF-1, in allogeneic (129) and syngeneic (B6) recipients. Strikingly, p45 -/- CD8 cells were greatly outcompeted by WT CD8 cells in 129 recipients in all tissues and at all times post-transplant, among both Tet H60+ and Tet H60- cells. In contrast, in B6 recipients, WT and p45 -/- cells remained evenly matched, suggesting that full-length TCF-1 isoforms are dispensable for lymphopenia-induced T cell expansion. Further, p45 -/- cells were also not disadvantaged when adoptively transferred into B6 mice and acutely challenged with H60 antigen by vaccination. Together these data suggest a model wherein TCF-1 hi progenitor like T cells are seeded in GVHD target organs where they may serve as a key local source for GVHD effectors, and moreover, full-length TCF-1 is itself critical for alloreactive T cell fitness in GVH responses.

Disclosures Shlomchik: BlueSphere Bio: Current Employment, Current holder of stock options in a privately-held company, Membership on an entity's Board of Directors or advisory committees, Patents & Royalties, Research Funding; Orca Bio: Consultancy, Current holder of stock options in a privately-held company.

https://doi.org/10.1182/blood-2023-190889